0040-4020(94)E0359-2

# Stereocontrolled Synthesis of $\beta$ -Hydroxyphenylalanine and $\beta$ -Hydroxytyrosine Derivatives

Christopher J. Easton,\* Craig A. Hutton, Peter D. Roselt and Edward R. T. Tiekink

Department of Chemistry, University of Adelaide, Adelaide, South Australia 5005, Australia

Abstract: Side-chain bromination of N-phthaloyl-(S)-phenylalanine and tyrosine derivatives, followed by treatment of the product bromides with silver nitrate in aqueous actone, affords the corresponding (2S,3R)- $\beta$ -hydroxy- $\alpha$ -amino acids, enantiospecifically and diastereoselectively. The diastereoselectivity depends on the carboxyl protecting group. *tert*-Butyl esters display greater stereoselectivity than the corresponding methyl esters, presumably as a result of a steric effect, while N-tert-butylamides react diastereospecifically due to a combination of steric and electronic effects.

### **INTRODUCTION**

 $\beta$ -Hydroxyphenylalanine 1 and  $\beta$ -hydroxytyrosine 2 are important naturally occurring amino acids. They have been implicated as precursors in the biosynthesis of the hypertensive agents adrenalin and noradrenalin,<sup>1</sup> and of the antibiotic chloramphenicol,<sup>2</sup> and as components of peptidases<sup>3</sup> and esterases.<sup>4</sup> They have also been identified as components of biologically active cyclic peptides. As examples, vancomycin contains two residues of  $\beta$ -hydroxytyrosine 2, one with the (2S,3R)-stereochemistry and the other with the (2R,3R)-stereochemistry,<sup>5</sup> lysobactin<sup>6</sup> contains  $\beta$ -hydroxyphenylalanine 1 of the (2S,3R)-stereochemistry, while phomopsin A<sup>7</sup> contains  $\beta$ -hydroxyphenylalanine 1 and bouvardin<sup>8</sup> contains a residue of  $\beta$ hydroxytyrosine 2, each with the (2S,3S)-stereochemistry. Hydroxy amino acids are also of interest as enzyme inhibitors.<sup>9-11</sup> For example,  $\beta$ -hydroxyphenylalanine 1 has been shown to inhibit Neisseria gonorrhoeae bacterial strains<sup>10</sup> and the lactose operon in Escherichia coli.<sup>11</sup>



As a consequence of their biochemical activity, there is considerable interest in efficient routes for the stereocontrolled synthesis of the hydroxy amino acids 1 and 2, and related compounds. Many asymmetric syntheses of  $\beta$ -hydroxy amino acids *via* condensation of glycine equivalents with aldehydes have been reported.<sup>12-15</sup> General and versatile methods have been developed by Schöllkopf *et al.*,<sup>13</sup> by Seebach and coworkers<sup>14</sup> and by Evans and Weber.<sup>15</sup> Although these procedures give products of high enantiomeric excess,





there remains a strong demand for compounds which are enantiomerically pure. A method for the enantiospecific synthesis of  $\beta$ -hydroxy- $\alpha$ -amino acids from D-glucose has been reported by Rao *et al.*,<sup>16</sup> but the procedure involves many steps. Shimamoto and Ohfune<sup>17</sup> have developed a novel synthesis of (2S,3R)- $\beta$ -hydroxy-O-methyltyrosine 4, by direct benzylic oxidation of the *N*-tert-butoxycarbonyltyrosinol derivative 3 (Scheme 1). This procedure is limited by its lack of generality, however, with attempts to synthesize the (2S,3R)-isomer of  $\beta$ -hydroxyphenylalanine 1 via an analogous pathway being unsuccessful.<sup>17</sup>

Recently we reported<sup>18</sup> preliminary details of a complementary method for the enantiospecific and diastereoselective synthesis of  $\beta$ -hydroxy- $\alpha$ -amino acids, which involved direct side-chain bromination of amino acid derivatives<sup>19,20</sup> followed by treatment of the product bromides with aqueous silver nitrate. For example, the phenylalanine derivative 5a gave a 1:1 mixture of the diastereomeric bromides 6a and 7a, and that mixture gave a 5:1 mixture of the (2S,3R)-hydroxyphenylalanine derivative 8a and the (2S,3S)-diastereomer 9a. We now report our investigation of the origin of the stereoselectivity of the hydrolysis, together with full details of our earlier work. We also describe an unusual substituent effect which results in the enantiospecific and diastereospecific synthesis of derivatives of  $\beta$ -hydroxyphenylalanine and  $\beta$ -hydroxytyrosine.

## **RESULTS AND DISCUSSION**

As previously reported,<sup>20,21</sup> treatment of the phenylalaninamide 5c with N-bromosuccinimide gave a 1:1 mixture of the diastereomeric bromides 6c and 7c, which were separated by fractional crystallization. The absolute stereochemistry of the bromides 6c and 7c is predetermined by that of the phenylalanine derivative 5c, while their relative stereochemistry has been determined previously, for each of the corresponding racemates, through their *anti*-elimination to dehydro amino acid derivatives on treatment with potassium fluoride.<sup>21</sup>



Bromination of the ester 5a gave a 1:1 mixture of the bromides 6a and 7a, which were separated by fractional crystallization. The relative stereochemistry of each of the bromo esters 6a and 7a was determined using the procedure reported<sup>21</sup> for determining the stereochemistry of the corresponding bromo amides 6c and 7c. Treatment of the bromo ester 6a with potassium fluoride produced the (Z)-dehydrophenylalanine derivative 10 in 88% yield, but none of the (E)-isomer 11. The structure and stereochemistry of the (Z)-alkene 10 was confirmed through X-ray crystallographic analysis.<sup>22</sup> Treatment of the bromide 7a with potassium fluoride gave a 2:1 mixture of the (E)-dehydrophenylalanine derivative 11 and the (Z)-alkene 10. The stereochemistry of the (E)-alkene 11 was confirmed by comparison of its <sup>1</sup>H NMR spectrum with that of the (Z)-isomer 10, where the signal due to the vinylic proton of the (E)-alkene 11 occurred 0.9 ppm upfield from that of the (Z)-isomer 10.<sup>21,23</sup> Presumably the bromides 6a and 7a undergo selective *anti*-elimination, on which basis the bromide 6a that gave only the (Z)-alkene 10 can be assigned the (2R,3R)-stereochemistry. Note that the Cahn-Ingold-Prelog designation at the  $\alpha$ -carbon of the bromides 6a and 7a is reversed by comparison with that of the precursor 5a, due to the change in the priority of substituents.

The tert-butyl ester 5b and the tyrosine derivatives 12a and 12b reacted with N-bromosuccinimide to give the corresponding benzylic bromides 6b and 7b, 13a and 14a, and 13b and 14b. The diastereomeric bromophenylalanine derivatives 6b and 7b were separated by fractional crystallization. Although the diastereomeric tyrosine derivatives 13a and 14a, and 13b and 14b, were not completely separated, due to their instability, samples enriched in each stereoisomer were obtained by chromatography. The relative stereochemistry of the bromides 6b, 7b, 13a,b and 14a,b was determined by comparison of their <sup>1</sup>H NMR spectra with those of the bromides 6a,c and 7a,c (Table 1), which follow a general trend. The signals corresponding to the carboxyl protecting groups occur at lower chemical shift for the (2R,3R)-diastereomers 6a-c and 13a,b than for the corresponding (2R,3S)-diastereomers 7a-c and 14a,b. Also, the (2R,3R)-stereoisomers 6a-c and 13a,b exhibit the  $\beta$ -proton signal at higher chemical shift, the  $\alpha$ -proton signal at lower chemical shift, and a larger coupling constant between the  $\alpha$ - and  $\beta$ -protons, than for the corresponding (2R,3S)-diastereomers 7a-c and 14a,b. The cause of these effects may be explained by considering, as an example, the preferred conformation of each of the bromides 6a and 7a (Figure 1). It can be seen that with the



(2R,3R)-isomer 6a, the phenyl group is situated close to the ester moiety, and the shielding effect of the phenyl group may explain the lower chemical shift of the signal due to the ester group protons. In the case of the (2R,3S)-bromide 7a,  $\pi,\pi$ -stacking between the phthalimido and phenyl groups would cause rotation about the C $\alpha$ -C $\beta$  bond, such that the dihedral angle between the  $\alpha$ - and  $\beta$ -protons would be less than 180°, thereby explaining the lower coupling constant observed between the  $\alpha$ - and  $\beta$ -protons for the (2R,3S)-diastereomer 7a than for the (2R,3R)-diastereomer 6a.<sup>24</sup>

The reactions of the amino acid derivatives **5a-c** and **12a,b** to give the corresponding bromides **6a-c** and **7a-c**, and **13a,b** and **14a,b** occur without discernible asymmetric induction, presumably as a result of the low activation energy for halogen transfer to the intermediate benzylic radicals.<sup>25</sup>

Treatment of the (2R,3R)-bromophenylalanine derivative 6a with silver nitrate in acetone/water<sup>26</sup> gave a

Compound	Stereochemistry	Chemical shift (0) / J (Hz)					
		α-Η	β-Н	$J_{\alpha,\beta}$	Carboxyl protecting group	OAc	
6a	2R,3R	5.42	5.95	11.2	3.50	-	
7a	2R,3S	5.55 <sup>a</sup>	5.92 <sup>a</sup>	10.5	3.80	-	
6b	2R,3R	5.47	6.06	11.4	1.16	-	
7b	2R,3S	5.49	5.85	10.4	1.48	-	
6c	2R,3R	5.28	6.25	11.8	1.03	-	
7c	2R,3S	5.32	6.04	11.4	1.43	-	
13a	2R,3R	5.46	6.03	11.2	3.56	2.30	
14a	2R,3S	5.56	5.93	10.4	3.81	2.19	
13b	2R,3R	5.17	6.22	11.8	1.05	2.31	
14b	2R,3S	5.30	6.08	11.5	1.38	2.20	

Table 1. <sup>1</sup>H NMR Spectral Data of the Bromides 6a-c, 7a-c, 13a,b and 14a,b.

<sup>a</sup>Assigned with the aid of proton-carbon heterocorrelation NMR spectroscopy.



Figure 1. Preferred Conformation of each of the Bromides 6a and 7a.

2:1 mixture of the (2S,3R)- $\beta$ -hydroxyphenylalanine derivative **8a** and the (2S,3S)-diastereomer **9a**. The ratio of the diastereomers **8a** and **9a** was determined through analysis of the <sup>1</sup>H NMR spectrum of the crude reaction mixture. Recrystallization of the crude product gave the (2S,3R)- $\beta$ -hydroxyphenylalanine derivative **8a**, in 75% yield, the stereochemistry of which was determined using X-ray crystallographic analysis.<sup>27</sup> Purification of the (2S,3S)-alcohol **9a** in the recrystallization mother liquor was achieved using HPLC. Treatment of the (2R,3S)-bromide **7a** under the same conditions as described for the hydrolysis of the (2R,3R)-bromide **6a** gave only the (2S,3R)-hydroxyphenylalanine derivative **8a**, in 93% yield.

The <sup>1</sup>H NMR spectra of the hydroxyphenylalanine derivatives **8a** and **9a** follow a general trend displayed in the spectra of the alcohols **8a-c**, **9a-c**, **15a,c** and **16a,c**, described herein (Table 2). The chemical shifts of the signals due to the  $\alpha$ - and  $\beta$ -protons and the carboxyl protecting groups of the (2*S*,3*R*)isomers **8a-c** and **15a,c** are significantly higher than those of the corresponding (2*S*,3*S*)-isomers **9a-c** and **16a,c**. Also, the (2*S*,3*R*)-isomers **8a-c** and **15a,c** each exhibit a smaller coupling constant between their  $\alpha$ and  $\beta$ -protons than is observed for the corresponding (2*S*,3*S*)-isomers **9a-c** and **16a,c**. The correlation of the coupling constants with the stereochemistry may be attributed to the alcohols **8a-c**, **9a-c**, **15a,c** and **16a,c** adopting hydrogen-bonded conformations, as shown in Figure 2 for the phenylalanine derivatives **8a** and **9a**. The dihedral angle between the  $\alpha$ - and  $\beta$ -protons of the (2*S*,3*R*)-isomers **8a-c** and **15a,c** in these conformations would be approximately 60°, whereas that angle would be approximately 180° for the (2*S*,3*S*)isomers **8a-c** and **15a,c** would be smaller than that observed for each of the corresponding (2*S*,3*S*)-isomers **9a-c** and **15a,c**.



Figure 2. Hydrogen-bonded Conformation of each of the Alcohols 8a and 9a.

Compound	Stereochemistry	Chemical shift ( $\delta$ ) / J (Hz)					
		α-Η	β-Н	$J_{\alpha,\beta}$	Carboxyl protecting group	OAc	
8a	2 <i>S</i> ,3 <i>R</i>	5.51	5.71	4.6	3.86	-	
9a	2 <i>S</i> ,3 <i>S</i>	5.02	5.52	8.4	3.79		
8b	2 <i>S</i> ,3 <i>R</i>	5.44	5.67	4.6	1.52	-	
9b	2 <i>S</i> ,3 <i>S</i>	4.95	5.49	8.0	1.28		
8c	2S,3R	5.11	5.63	6.2	1.30	-	
9c	2S,3S	4.61	5.39	8.3	1.15		
15a	2S,3R	5.48	5.67	5.0	3.78	2.19	
16a	2S,3S	5.02	5.54	8.2	3.74	2.17	
15b	2S,3R	5.08	5.63	6.3	1.31	2.26	
15c	2 <i>S</i> ,3 <i>R</i>	5.44	5.64	4.8	3.85	-	
16c	2 <i>S</i> ,3 <i>S</i>	4.97	5.49	8.6	3.78		
15d	2S,3R	5.11	5.62	6.9	1.27	-	

Table 2. <sup>1</sup>H NMR Spectral Data of the Alcohols 8a-c, 9a-c, 15a-d and 16a,c.

The stereochemical courses of the substitution reactions of the bromides 6a and 7a indicate that they occur via different mechanisms. It appears that the (2R, 3R)-bromide **6a** reacts via the carbocation **17a**, while the (2R.3S)-bromide 7a undergoes an SN2 reaction. This variation in the mechanisms and the stereochemical courses of the reactions can be rationalized by considering the preferred conformation of each of the bromides **6a** and **7a** (Figure 1). In the preferred conformation of the (2R, 3R)-bromide **6a**, the phenyl and  $\beta$ -hydrogen substituents are already in the orientation required to form the most stable conformation of the carbocation 17a (Figure 3). This orientation also allows for delocalization of the developing positive charge by the ester carbonyl group, as the ester moiety is situated in the plane of the developing unoccupied orbital.  $S_N 2$ substitution is disfavored for this conformation of the bromide 6a, as the ester moiety blocks attack of water from behind the carbon-bromine bond. For these reasons, the carbocation 17a forms, with subsequent nucleophilic attack of water preferentially from the less hindered face, opposite the ester group, giving rise to the alcohols 8a and 9a in a 2:1 ratio. In its preferred conformation, the (2R,3S)-bromide 7a is not aligned to lead directly to the most stable conformation of the carbocation 17a. In fact, the bromide 7a can only react to give the carbocation 17a either initially in a less stable conformation or without delocalization of the developing positive charge by the ester carbonyl group. These processes are energetically less favourable than the reaction of the bromide 6a to give the carbocation 17a and, instead, the bromide 7a reacts via the SN2 pathway with inversion of configuration.

Subsequent hydrolyses were performed using 1:1 mixtures of the diastereomeric bromides **6b**,c and **7b**,c, and **13a**,b and **14a**,b, thereby avoiding their separation. For comparison, reaction of a 1:1 mixture of the bromides **6a** and **7a** gave the alcohols **8a** and **9a** in the expected 5:1 ratio. Initially, the reaction of the *tert*butyl esters **6b** and **7b** was examined, because it was envisaged that the more bulky ester group would increase the stereoselectivity of the reaction. In the event, the alcohols **8b** and **9b** were formed in an 8:1 ratio, as



Figure 3. Direct Formation of the Most Stable Conformation of the Carbocation 17a from the Preferred Conformation of the Bromide 6a.

determined through analysis of the <sup>1</sup>H NMR spectrum of the crude reaction mixture. The (2S,3R)-isomer 8b was isolated in 81% yield, after recrystallization of the crude product. Presumably, by analogy with the reaction of the methyl ester 7a, the reaction of the (2R,3S)-bromide 7b occurs with complete inversion of stereochemistry. The reaction of the (2R,3R)-bromide 6b must therefore occur to give an approximately 3.5:1 ratio of the alcohols 8a and 9a. The increased selectivity of the reaction of the *tert*-butyl ester 6b, compared to that of the methyl ester 6a, can be attributed to the relative extent of the steric effects of the respective ester moleties, blocking one face of each of the corresponding intermediate carbocations 17b and 17a.

Treatment of a mixture of the phenylalaninamides 6c and 7c with silver nitrate in acetone/water gave only the (2S,3R)-alcohol 8c, in 93% yield. In order to determine that none of the stereoisomer 9c was produced in this reaction, an authentic sample was prepared. Oxidation of the alcohol 8c with Jones reagent<sup>28</sup> gave the ketone 18, which was reduced with sodium borohydride in ethanol to give a 1.2:1 mixture of the alcohols 8c and 9c. It is possible that some racemization occurred during this oxidation-reduction sequence, but the absolute stereochemistry of the alcohol 9c is of no consequence in determining the diastereoselectivity of the reaction of the bromides 6c and 7c. When the <sup>1</sup>H NMR spectrum of the crude reaction mixture obtained from the hydrolysis of the bromides 6c and 7c was compared with that of the alcohol 9c, the former showed no signal corresponding to the *tert*-butyl group of the alcohol 9c, even though the <sup>13</sup>C satellite peaks of that signal for the alcohol 8c ( $J_{CH} = 126.7$  Hz) were clearly visible, with a signal to noise ratio of greater than 10:1. On that basis the hydrolysis of the mixture of the bromides 6c and 7c gave the (2*S*,3*R*)-alcohol 8c in >99.9% diastereomeric excess.

The diastereoselectivity of the hydrolysis of the mixture of the bromo amides 6c and 7c is at least 100fold greater than that for the reaction of the bromo esters 6b and 7b. This may be attributed to the relative





Figure 4. Stabilization of the Carbocation 17c by the Amido Substituent.

extent of conformational preference of the carbocations 17b and 17c. A carbonyl oxygen of an amido group is approximately six orders of magnitude more basic than that of an ester,<sup>29</sup> therefore the extent of interaction of the amido substituent with the unoccupied orbital of the carbocation 17c (Figure 4) is likely to be much greater than the analogous interaction of the ester group in the carbocation 17b. This will result in a greater conformational preference of the carbocation 17c and a greater tendency for approach of the nucleophile to the face of the carbocation 17c opposite to that through which the amido group stabilization occurs.

A substituent effect analogous to that observed in the reactions of the phenylalanine derivatives 6a,c and 7a,c was also observed with the corresponding tyrosine derivatives 13a,b and 14a,b. Treatment of a 1:1 mixture of the diastereomeric esters 13a and 14a under the standard hydrolysis conditions gave a 6:1 mixture of the alcohols 15a and 16a, in 86% yield. The hydroxytyrosine derivatives 15a and 16a were unstable and deacetylated slowly on exposure to moisture. Hence, the crude product was completely deacetylated by treatment with a catalytic amount of *p*-toluenesulfonic acid in methanol, to give a 6:1 ratio of the alcohols 15c and 16c, from which the (2S,3R)-isomer 15c was isolated, in 72% yield, by recrystallization. HPLC of the mother liquor afforded a pure sample of the (2S,3S)-alcohol 16c. Reaction of a 1:1 mixture of the tyrosinamides 13b and 14b gave only the (2S,3R)-stereoisomer 15b, in 88% yield, which on deacetylation afforded only the alcohol 15d, in 93% yield. Within the limits of detection using <sup>1</sup>H NMR spectroscopy, the reaction of the mixture of the bromides 13b and 14b was diastereospecific, as there was no evidence of formation of either of the alcohols 16b or 16d.

The reactions of the amino acid derivatives 5a-c and 12a,b with N-bromosuccinimide, and the subsequent treatment of the product bromides 6a-c, 7a-c, 13a,b and 14a,b with silver nitrate in aqueous acetone, described above, exemplify an efficient route for the stereocontrolled synthesis of hydroxy amino acid derivatives. The alcohols 8a and 8c were also used to prepare the (2S,3R)-stereoisomer of the corresponding free amino acid 1. Treatment of the ester 8a with hydrazine to remove the phthaloyl group, <sup>30</sup> followed by acid-catalysed hydrolysis of the ester moiety, gave the hydrochloride salt of the (2S,3R)-stereoisomer of  $\beta$ -hydroxyphenylalanine 1. Alternatively, the salt was obtained by treatment of the ester 8a with a 2:1 mixture of 6N hydrochloric acid and glacial acetic acid, at reflux. In each case the free amino acid was prepared from the hydrochloric acid and glacial acetic acid, at reflux, followed by crystallization of the crude product from aniline in ethanol. Also gave the corresponding free amino acid, in 78% yield. The relative and absolute stereochemistry of samples of the free amino acid, prepared from the amino acid derivatives 8a and 8c, was established by comparison of their <sup>1</sup>H NMR spectra<sup>31</sup> and optical rotation properties<sup>32</sup> with literature data. This confirmed that the syntheses of the (2S,3R)-diastereomer of the alcohol 1 from (S)-phenylalanine occurred with

retention of chirality at the  $\alpha$ -position. It follows that identical procedures could be used in the elaboration of (R)-amino acids. Thus, the procedures described above are suitable for the enantiospecific and diastereospecific synthesis of  $\beta$ -hydroxy- $\alpha$ -amino acids and their derivatives, particularly the (2S,3R)- and (2R,3S)-stereoisomers.

## **EXPERIMENTAL**

General. General experimental details have been reported previously.<sup>20</sup> <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded at 300 and 75.5 MHz, respectively, in deuteriochloroform unless otherwise stated. Infrared spectra were recorded as solutions in dichloromethane. Organic solutions were dried by stirring over anhydrous magnesium sulfate.

(S)-Phenylalanine and (S)-tyrosine were purchased from Sigma Chemical Co., and used to prepare the amino acid derivatives 5a-c and 12a,b, respectively, using standard procedures.<sup>20,21,33</sup>

Bromination of the Amino Acid Derivatives 5a-c and 12a,b. Reactions of the amino acid derivatives 5a-c and 12a,b with N-bromosuccinimide were carried out as described previously<sup>20,21</sup> for the bromination of the racemate of the amide 5c, except that the reactions of the esters 5a,b and 12a were performed in carbon tetrachloride, instead of the 3:1 mixture of carbon tetrachloride/dichloromethane that was used with the amides 5c and 12b. The reactions gave 1:1 mixtures of the diastereomeric bromides 6a-c and 7a-c, and 13a,b and 14a,b, which were separated by fractional crystallization in the cases of the phenylalanine derivatives 6a-c and 7a-c, from hexane/dichloromethane in the cases of 6a,b and 7a,b, and from hexane/2-propanol in the case of 6c and 7c.

(2R,3R)-3-Bromo-N-phthaloylphenylalanine Methyl Ester (6a): 42%; mp 142-143 °C;  $v_{max}$  1778, 1755, 1709, 708 cm<sup>-1</sup>; MS (EI) *m/e* 389 and 387 (M<sup>+</sup>, 6%), 330 (8), 328 (8), 308 (65), 307 (18), 276 (89), 249 (92), 248 (100), 242 (32), 240 (32), 218 (59), 190 (46), 162 (39), 130 (22), 105 (46), 103 (28), 77 (17). Anal. Calcd for C<sub>18</sub>H<sub>14</sub>BrNO<sub>4</sub>: C, 55.8; H, 3.7; N, 3.6. Found: C, 55.7; H, 3.7; N, 3.6.

(2R,3S)-3-Bromo-N-phthaloylphenylalanine Methyl Ester (7a): 40%; mp 121-122 °C; <sup>13</sup>C NMR  $\delta$  47.6, 53.1, 57.0, 123.6, 128.1, 128.6, 128.9, 130.9, 134.3, 137.1, 166.3, 167.2;  $\nu_{max}$  1774, 1758, 1718, 727 cm<sup>-1</sup>; MS (EI) *m/e* 389 and 387 (M<sup>+</sup>, 2%), 330 (5), 328 (5), 308 (32), 276 (59), 249 (79), 248 (85), 242 (24), 240 (24), 218 (56), 190 (62), 169 (11), 167 (11), 161 (26), 130 (33), 104 (82), 102 (64), 76 (100). Anal. Calcd for C<sub>18</sub>H<sub>14</sub>BrNO<sub>4</sub>: C, 55.8; H, 3.7; N, 3.6. Found: C, 56.2; H, 3.7; N, 3.7.

(2*R*,3*R*)-3-Bromo-*N*-phthaloylphenylalanine *tert*-Butyl Ester (6b): 41%; mp 152-153 °C; v<sub>max</sub> 1780, 1738, 1720, 1420, 1390, 1050, 840, 710 cm<sup>-1</sup>; MS (EI) *m/e* 431 and 429 (M<sup>+</sup>, 0.02%), 375 (9), 373 (9), 330 (11), 328 (11), 294 (4), 276 (2), 249 (93), 248 (94), 232 (15), 220 (10), 204 (44), 165 (7), 130 (8), 104 (38), 102 (32), 76 (36), 57 (100).

(2R,3S)-3-Bromo-N-phthaloylphenylalanine *tert*-Butyl Ester (7b): 39%; mp 118-119 °C; v<sub>max</sub> 1780, 1748, 1726, 1390, 715 cm<sup>-1</sup>; MS (EI) *m/e* 431 and 429 (M<sup>+</sup>, 0.01%), 375 (7), 373 (7), 330 (10), 328 (10), 294 (6), 276 (3), 249 (75), 248 (100), 232 (11), 220 (8), 204 (28), 165 (5), 130 (6), 104 (27), 102 (25), 76 (26), 57 (66).

(2R,3R)-3-Bromo-N-tert-butyl-N<sup> $\alpha$ </sup>-phthaloylphenylalaninamide (6c): 43%; mp 208-209 °C;  $\nu_{max}$  3350, 1700, 1530, 1450, 1365, 710 cm<sup>-1</sup>; MS (EI) *m/e* 430 and 428 (M<sup>+</sup>, 5%), 415 (0.2), 413 (0.2), 375 (0.6), 373 (0.6), 330 (2), 328 (2), 249 (100), 232 (6), 220 (3), 204 (5), 165 (2), 130 (3), 104 (8), 102 (7), 76 (6). Anal. Calcd for C<sub>21</sub>H<sub>21</sub>BrN<sub>2</sub>O<sub>3</sub>: C, 58.8; H, 4.9; N, 6.5. Found: C, 58.6; H, 5.0; N, 6.8.

(2R,3S)-3-Bromo-N-tert-butyl-N<sup> $\alpha$ </sup>-phthaloylphenylalaninamide (7c): 41%; mp 188-191 °C; v<sub>max</sub> 3375, 1775, 1705, 1530, 1380, 720 cm<sup>-1</sup>; MS (EI) *m/e* 430 and 428 (M<sup>+</sup>, 1%), 415 (0.1), 413 (0.1), 375 (0.2), 373 (0.2), 330 (1), 328 (1), 249 (100), 232 (10), 220 (3), 204 (4), 165 (2), 130 (3), 104 (12), 102 (10), 76 (13). Anal. Calcd for C<sub>21</sub>H<sub>21</sub>BrN<sub>2</sub>O<sub>3</sub>: C, 58.8; H, 4.9; N, 6.5. Found: C, 58.8; H, 4.9; N, 6.6.

(2R,3R)-O-Acetyl-3-bromo-N-phthaloyltyrosine Methyl Ester (13a) and (2R,3S)-O-Acetyl-3-bromo-N-phthaloyltyrosine Methyl Ester (14a): 1:1 mixture; 97%; MS (EI) *m/e* 447 and 445 (M<sup>+</sup>, 0.3%), 405 (0.3), 403 (0.3), 388 (0.6), 386 (0.6), 366 (8), 334 (17), 324 (5), 306 (5), 292 (16), 264 (100), 218 (12), 187 (7), 163 (9), 147 (10), 138 (14), 121 (35), 104 (22), 86 (38), 76 (19); MS (EI) *m/e* 445.018 (M<sup>+</sup>). Calc. for C<sub>20</sub>H<sub>16</sub>BrNO<sub>6</sub>: *m/e* 445.016.

(2R,3R)-O-Acetyl-3-bromo-N-tert-butyl-N<sup>\alpha</sup>-phthaloyltyrosinamide (13b) and (2R,3S)-O-Acetyl-3-bromo-N-tert-butyl-N<sup>\alpha</sup>-phthaloyltyrosinamide (14b): 1:1 mixture; 97%; v<sub>max</sub> 1780, 1726, 1675, 1608, 1394, 1160, 720 cm<sup>-1</sup>; MS (EI) *m/e* 488 and 486 (M<sup>+</sup>, 2%), 446 (1), 444 (1), 403 (14), 388 (1), 386 (1), 361 (10), 308 (31), 265 (100), 248 (5), 121 (11), 118 (10), 104 (14), 76 (9); MS (EI) *m/e* 486.080 (M<sup>+</sup>). Calc. for C<sub>23</sub>H<sub>23</sub>BrN<sub>2</sub>O<sub>5</sub>: *m/e* 486.079.

(2S,3R)-3-Hydroxy-N-phthaloylphenylalanine Methyl Ester (8a) and (2S,3S)-3-Hydroxy-N-phthaloylphenylalanine Methyl Ester (9a). To a solution of a 1:1 mixture of the bromides 6a and 7a (1.0 g, 2.6 mmol) in acetone (40 ml) was added a solution of silver nitrate (0.66 g, 3.9 mmol) in water (40 ml), and the resultant mixture was stirred in the dark at rt for 24 h, then it was filtered and the filtrate was concentrated under reduced pressure. The residue was extracted with dichloromethane, and the organic solution was dried and then concentrated under reduced pressure, to give a 5:1 mixture of the alcohols 8a and 9a (0.77 g, 92%), as determined from the <sup>1</sup>H NMR spectrum. Recrystallization of the mixture from hexane/dichloromethane gave the alcohol 8a as colourless crystals (0.63 g, 75%): mp 185-186 °C;  $[\alpha]_D^{16}$ -67.0° (c, 0.6 in EtOH);  $v_{max}$  3580, 3420, 1780, 1745, 1715, 1390, 1215, 1020, 720 cm<sup>-1</sup>; MS (FAB) *m/e* 326 (M+H<sup>+</sup>, 77%), 308 (100), 248 (54), 219 (19), 160 (27), 149 (23), 131 (21), 105 (48), 104 (33), 91 (42). Anal. Calcd for C<sub>18</sub>H<sub>15</sub>NO<sub>5</sub>: C, 66.5; H, 4.7; N, 4.3. Found: C, 66.7; H, 4.7; N, 4.4.

The structure of the alcohol 8a was confirmed through X-ray crystallographic analysis.27

Reactions of the bromides 6b,c and 7b,c, and 13a,b and 14a,b, with silver nitrate in aqueous acetone. These reactions were performed using the procedure described above for the reaction of the mixture of the bromides 6a and 7a.

(2S,3R)-3-Hydroxy-N-phthaloylphenylalanine tert-Butyl Ester (8b) and (2S,3S)-3-Hydroxy-N-phthaloylphenylalanine tert-Butyl Ester (9b). Reaction of a 1:1 mixture of the bromides 6b and 7b gave an 8:1 mixture of the alcohols 8b and 9b. The ratio of the diastereomers 8b and 9b was determined from the <sup>1</sup>H NMR spectrum of the mixture. Recrystallization of the mixture from hexane/dichloromethane gave the alcohol 8b as colourless crystals: 81%; mp 125-127 °C; v<sub>max</sub> 3440, 1785, 1738, 1712, 1604, 1552, 1392, 1194, 1108, 844, 720 cm<sup>-1</sup>; MS (FAB) m/e 368 (M+H<sup>+</sup>, 8%), 312 (44), 294 (55), 276 (23), 266 (10), 250 (100), 232 (11), 205 (13), 160 (19), 105 (21), 91 (20), 87 (25), 57 (93). Anal. Calcd for C<sub>21</sub>H<sub>21</sub>NO<sub>5</sub>: C, 68.7; H, 5.8; N, 3.8. Found: C, 68.7; H, 5.9; N, 3.8.

(2S,3R)-3-Hydroxy-*N*-tert-butyl-N<sup> $\alpha$ </sup>-phthaloylphenylalaninamide (8c). Reaction of a 1:1 mixture of the bromides 6c and 7c gave the alcohol 8c, as colourless crystals from hexane/dichloromethane: 93%; mp 195-197 °C; v<sub>max</sub> 3352, 3275, 1778, 1704, 1644, 717 cm<sup>-1</sup>; MS (FAB) *m/e* 367 (M+H<sup>+</sup>, 68%), 307 (3), 289 (2), 266 (4), 260 (18), 250 (100), 232 (7), 187 (10), 160 (17), 154 (24), 136 (18), 107 (8), 105 (6), 77 (6). Anal. Calcd for C<sub>21</sub>H<sub>22</sub>N<sub>2</sub>O<sub>4</sub>: C, 68.8; H, 6.1; N, 7.7. Found: C, 68.8; H, 6.0; N, 7.7.

(2S,3R)-3-Hydroxy-N-phthaloyltyrosine Methyl Ester (15c) and (2S,3S)-3-Hydroxy-N-phthaloyltyrosine Methyl Ester (16c). Reaction of a 1:1 mixture of the bromides 13a and 14a gave an 86% yield of a 6:1 mixture of the alcohols 15a and 16a:  $v_{max}$  3580, 3410, 1775, 1752, 1712, 1552, 1392, 1184, 1118, 852, 710 cm<sup>-1</sup>; MS (FAB) *m/e* 384 (M+H<sup>+</sup>, 13%), 366 (79), 334 (100), 324 (7), 306 (35), 292 (38), 264 (58), 219 (34), 187 (23), 160 (15), 154 (37), 136 (33), 107 (17), 105 (13), 104 (12), 89 (17), 77 (19); MS *m/e* 365.090 (M<sup>+</sup>). Calc. for C<sub>20</sub>H<sub>15</sub>NO<sub>6</sub>: *m/e* 365.090. The ratio of the diastereomers 15a and 16a was determined from the <sup>1</sup>H NMR spectrum of the mixture.

The acetate moieties of the alcohols 15a and 16a were found to hydrolyse slowly on exposure to moisture and the mixture was therefore deacetylated without further purification. A mixture of the acetates 15a and 16a (0.87 g, 2.27 mmol), *p*-toluenesulfonic acid monohydrate (100 mg, 0.5 mmol) and water (1 ml) in methanol (20 ml) was stirred at rt for 6 h, then it was concentrated under reduced pressure. The residue was partitioned between ethyl acetate and water, and the organic phase was separated and washed with 10% aqueous sodium carbonate, then it was dried and concentrated under reduced pressure, to give a 6:1 mixture of the alcohols 15c and 16c (0.74 g, 96%). The ratio of the diastereomers 15c and 16c was determined from the <sup>1</sup>H NMR spectrum of the mixture. Recrystallization of the mixture from hexane/ethyl acetate gave the alcohol 15c as a white solid (556 mg, 72%); mp 200-201 °C;  $[\alpha]_D^{16}$  -70.7° (c, 0.4 in EtOH); v<sub>max</sub> 3582, 3405, 1775, 1750, 1714, 1516, 1392, 1172, 850, 715 cm<sup>-1</sup>. MS (FAB) *m/e* 342 (M+H<sup>+</sup>, 2%), 324 (10), 307 (12), 292 (11), 289 (7), 264 (5), 232 (14), 231 (17), 219 (9), 154 (100), 137 (63), 136 (77), 107 (30), 105 (12), 89 (29), 77 (31). Anal. Calcd for C<sub>18</sub>H<sub>15</sub>NO<sub>6</sub>: C, 63.3; H, 4.4; N, 4.1. Found: C, 63.3; H, 4.4; N, 4.0.

(2S,3R)-3-Hydroxy-N-tert-butyl-N<sup> $\alpha$ </sup>-phthaloyltyrosinamide (15d). Reaction of a 1:1 mixture of the bromides 13b and 14b gave the alcohol 15b as colourless crystals; 88%; mp 202-204 °C; v<sub>max</sub> 3600, 3440, 3390, 1775, 1710, 1685, 1515, 1390, 1220, 860, 720 cm<sup>-1</sup>; MS (FAB) *m/e* 425 (M+H<sup>+</sup>, 43%), 308 (67), 266 (100), 248 (4), 187 (16), 160 (26), 154 (23), 136 (37), 107 (18), 105 (10), 89 (17), 77 (22); MS *m/e* 406.154 (M<sup>+</sup>-H<sub>2</sub>O). Calc. for C<sub>23</sub>H<sub>22</sub>N<sub>2</sub>O<sub>5</sub>: *m/e* 406.153.

The acetate **15b** was hydrolysed, as described above for the hydrolysis of the mixture of the acetates **15a** and **16a**, to give a 93% yield of the alcohol **15d**, as a colourless solid after recrystallization from hexane/chloroform: mp 214-215 °C;  $v_{max}$  3590, 3546, 3410, 1772, 1712, 1685, 1366, 855, 717 cm<sup>-1</sup>; MS (FAB) *m/e* 383 (M+H<sup>+</sup>, 18%), 307 (3), 289 (4), 267 (48), 266 (100), 260 (16), 187 (15), 160 (15), 154 (26), 136 (17), 107 (8), 105 (8), 91 (10), 77 (9). Anal. Calcd for C<sub>21</sub>H<sub>22</sub>N<sub>2</sub>O<sub>5</sub>: C, 66.0; H, 5.8; N, 7.3. Found: C, 65.7; H, 5.8; N, 7.2.

Isomerization of (2S,3R)-3-Hydroxy-*N*-tert-butyl-N<sup> $\alpha$ </sup>-phthaloylphenylalaninamide (8c). Jones reagent<sup>28</sup> (ca. 0.5 ml) was added dropwise to a vigorously stirred solution of the alcohol 8c (0.5 g, 1.37 mmol) in acetone (20 ml), until the red/brown colour persisted. The reaction mixture was stirred for a further 15 min at rt, then it was concentrated under reduced pressure. The residual oil was partitioned between dichloromethane and water. The organic layer was separated and washed with 10% aqueous sodium carbonate and with water, then it was dried and concentrated under reduced pressure. The residual oil was chromatographed on silica, eluting with ethyl acetate, to give the crude ketone 18, which recrystallized from hexane/ethyl acetate as a colourless solid (0.33 g, 67%); <sup>1</sup>H NMR  $\delta$  1.34 (9H, s), 6.04 (1H, s), 7.21-7.56 (5H, m), 7.73-7.90 (4H, m); MS (EI) *m/e* 364 (M<sup>+</sup>, 2%), 308 (1), 291 (2), 265 (40), 221 (6), 147 (72), 105 (43), 104 (87), 103 (58), 76 (100).

To a solution of the crude ketone 18 (100 mg, 0.27 mmol) in ethanol (5 ml) was added sodium borohydride (10 mg, 0.26 mmol). The mixture was stirred at rt for 10 min, then the reaction was quenched by the addition of dilute hydrochloric acid (*ca.* 1 ml). The mixture was concentrated under reduced pressure, and the residue was partitioned between dichloromethane and water. The organic layer was washed with 10% aqueous sodium carbonate and with water, then it was dried and concentrated under reduced pressure. Chromatography of the residual oil on silica gave a 1:1.2 mixture of (2S,3R)-3-hydroxy-N-tert-butyl- $N^{\alpha}$ -phthaloylphenylalaninamide (9c) and the diastereomer 8c (84 mg, 84%), as determined from the <sup>1</sup>H NMR spectrum.

Reaction of the Bromide 6a with Potassium Fluoride. Treatment of the bromide 6a with the 18-crown-6 complex of potassium fluoride in acetonitrile, as described previously<sup>21</sup> for the synthesis of (Z)-*N*-tert-butyl- $N^{\alpha}$ -phthaloyl-2,3-dehydrophenylalaninamide from the bromo amide 6c, afforded (Z)-*N*-phthaloyl-2,3-dehydrophenylalanine methyl ester (10) as colourless crystals: 88%; mp 136-137 °C; <sup>1</sup>H NMR  $\delta$  3.82 (3H, s), 7.26-7.42 (5H, m), 7.78-7.90 (4H, m), 8.12 (1H, s);  $v_{max}$  1780, 1720, 1640, 1600 cm<sup>-1</sup>; MS (EI) *m/e* 307 (M<sup>+</sup>, 100%), 279 (52), 248 (27), 247 (34). Anal. Calcd for C<sub>18</sub>H<sub>13</sub>NO4: C, 70.4; H, 4.3; N, 4.6. Found: C, 70.6; H, 4.2; N, 4.4.

The structure of the alkene 10 was confirmed through X-ray crystallographic analysis.<sup>22</sup>

Reaction of the Bromide 7a with Potassium Fluoride. Treatment of the bromide 7a with the 18-crown-6 complex of potassium fluoride in acetonitrile, as described for the reaction of the bromide 6a, afforded a 2:1 mixture of (*E*)-*N*-phthaloyl-2,3-dehydrophenylalanine methyl ester 11 and the (*Z*)-isomer 10, in 82% yield. Recrystallization of the mixture from hexane/ethyl acetate gave the (*Z*)-alkene 10 in 18% yield, while concentration of the recrystallization mother liquor afforded a 47% yield of a 5:1 mixture of the alkenes 11 and 10, as a clear oil;  $v_{max}$  1780, 1724, 1645, 1600, 1560 cm<sup>-1</sup>; MS (EI) *m/e* 307 (M<sup>+</sup>, 100%), 279 (49), 248 (12), 247 (27); <sup>1</sup>H NMR (11)  $\delta$  3.74 (3H, s), 7.23 (1H, s), 7.29-7.49 (5H, m), 7.80-7.93 (4H, m).

(2S,3R)-3-Hydroxyphenylalanine. A solution of the hydroxy ester 8a (0.25 g, 0.77 mmol) in a 2:1 mixture of 6N hydrochloric acid and acetic acid (10 ml) was heated at reflux for 5 h, then it was cooled and concentrated under reduced pressure. The residue was taken up in water (10 ml) and the suspension was filtered. The filtrate was concentrated under reduced pressure, and the residue was dissolved in a mixture of ethanol (7 ml), aniline (0.7 ml) and dichloromethane (10 ml). The mixture was stored at 0 °C for 16 h, and the precipitate that formed was collected by filtration, to give the (2S,3R)-isomer of the free amino acid 1, as a white powder (129 mg, 93%); mp 192-195 °C (lit.<sup>32</sup> 183-186 °C);  $[\alpha]_D^{16}$ -49.7±0.5° (c, 0.4 in 6N HCl) (lit.<sup>32</sup>

 $[\alpha]_D^{20}$  -50.2±2° (c, 2 in 6N HCl)); <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  3.95 (1H, d, J = 4.4 Hz), 5.29 (1H, d, J = 4.4 Hz), 7.47 (5H, m). Anal. Calcd for C<sub>9</sub>H<sub>11</sub>NO<sub>3</sub>: C, 59.7; H, 6.1; N, 7.7. Found: C, 59.9; H, 6.1; N, 7.8.

Treatment of the hydroxy amide 8c with hydrochloric acid and acetic acid, as described above for the hydrolysis of the hydroxy ester 8a, also gave the (2S,3R)-isomer of the free amino acid 1, in 78% yield.

ACKNOWLEDGEMENT. This work was supported by a grant from the Australian Research Council.

### REFERENCES

- 1. Rosenmund, K. W.; Dornsaft, H. Chem. Ber. 1919, 52, 1734.
- Hahn, F. E. In Antibiotics; Gottlieb, D.; Shaw, P. D. Eds., Springer-Verlag: Heidelberg, 1967, vol. 1, p. 308; Pongs, O. In Antibiotics; Hahn, F. E. Ed., Springer-Verlag: Heidelberg, 1979, vol. 5, part 1, p. 26.
- 3. Kraut, J. Ann. Rev. Biochem. 1977, 46, 331.
- Hammond, S. J.; Williamson, M. P.; Williams, D. H.; Boeck, L. D.; Marconi, G. G. J. Chem. Soc., Chem. Commun. 1982, 344; Hammond, S. J.; Williams, D. H.; Nielsen, R. V. J. Chem. Soc., Chem. Commun. 1983, 116; Lin, T.-S.; Kolattukudy, P. E. Eur. J. Biochem. 1980, 106, 341.
- 5. Williams, D. H. Acc. Chem. Res. 1984, 17, 364, and references cited therein.
- O'Sullivan, J.; McCullough, J. E.; Tymiak, A. A.; Kirsch, D. R.; Trejo, W. H.; Principe, P. A. J. Antibiot. 1988, 41, 1740; Tymiak, A. A.; McCormick, T. J.; and Unger, S. E. J. Org. Chem. 1989, 54, 1149.
- Culvenor, C. C. J.; Cockrum, P. A.; Edgar, J. A.; Frahn, J. L.; Gorst-Allman, C. P.; Jones, A. J.; Marasas, W. F. O.; Murray, K. E.; Smith, L. W.; Steyn, P. S.; Vleggaar, R.; Wessels, P. L. J. Chem. Soc., Chem. Commun. 1983, 1259; Mackay, M. F.; Van Donkelaar, A.; Culvenor, C. C. J. J. Chem. Soc., Chem. Commun. 1986, 1219.
- Jolad, S. D.; Hoffmann, J. J.; Torrance, S. J.; Wiedhopf, R. M.; Cole, J. R.; Arora, S. K.; Bates, R. B.; Gargiulo, R. L.; Kriek, G. R. J. Am. Chem. Soc. 1977, 99, 8040.
- 9. DiBello, I. C.; Dorling, P.; Fellows, L.; Winchester, B. FEBS Lett. 1984, 176, 61.
- 10. Hendry, A. T.; Dillon, J. R. Can. J. Microbiol. 1984, 30, 1319.
- 11. Janecek, J.; Jiresova, M.; Technikova, Z.; Spizek, J. Folia Microbiol. 1971, 16, 367.
- Guanti, G.; Banfi, L.; Narisano, E.; Scolastico, C. *Tetrahedron* 1988, 44, 3671.; Belokon, Y. N.; Bulychev, A. G.; Vitt, S. V.; Struchkov, Y. T.; Batsanov, A. S.; Timofeeva, T. V.; Tsyryapkin, V. A.; Ryzhov, M. G.; Lysova, L. A.; Bakhmutov, V. I.; Belikov, V. M. J. Am. Chem. Soc. 1985, 107, 4252; Casella, L.; Jommi, G.; Montanari, S.; Sisti, M. *Tetrahedron Lett.* 1988, 29, 2067; Ito, Y.; Sawamura, S.; Hayashi, T. J. Am. Chem. Soc. 1986, 108, 6405.
- Schöllkopf, U.; Groth, U.; Deng, C. Angew. Chem., Int. Ed. Engl. 1981, 20, 798; Schöllkopf, U.; Hartwig, W.; Pospischil, K.-H.; Kehne, H. Synthesis 1981, 966; Schöllkopf, U.; Nozulak, J.; Groth, U. Synthesis 1982, 868; Schöllkopf, U.; Groth, U. Angew. Chem., Int. Ed. Engl. 1981, 20, 977; Groth, U. Schöllkopf, U.; Synthesis 1983, 673; Scholkopf, U.; Tiller, T.; Bardenhagen, J. Tetrahedron 1988, 44, 5293.

- Seebach, D.; Imwinkelried, R.; Weber, T. Modern Synthetic Methods, Springer-Verlag: Heidelberg, 1986, vol. 4, 182.; Seebach, D.; Juaristi, E.; Miller, D. D.; Schickli, C.; Weber, T. Helv. Chim. Acta 1987, 70, 237.
- Evans, D. A.; Weber, A. E. J. Am. Chem. Soc. 1986, 108, 6757; Evans, D. A.; Weber, A. E. J. Am. Chem. Soc. 1987, 109, 7151.
- 16. Rao, A. V. R.; Yadav, J. S.; Chandraesekhar, S.; Rao, C. S. Tetrahedron Lett. 1989, 30, 6769.
- 17. Shimamoto, K.; Ohfune, Y. Tetrahedron Lett. 1988, 29, 5177.
- 18. Easton, C. J.; Hutton, C. A.; Tan, E. W.; Tiekink, E. R. T. Tetrahedron Lett. 1990, 31, 7059.
- 19. Easton, C. J.; Tan, E. W.; Hay, M. P. J. Chem. Soc., Chem. Commun. 1989, 385.
- 20. Easton, C. J.; Hutton, C. A.; Rositano, G.; Tan, E. W. J. Org. Chem. 1991, 56, 5614.
- 21. Easton, C. J.; Hutton, C. A.; Roselt, P. D.; Tiekink, E. R. T. Aust. J. Chem. 1991, 44, 687.
- 22. Easton, C. J.; Hutton, C. A.; Roselt, P. D.; Tiekink, E. R. T. Z. Krist., in press.
- 23. Srinivasan, A.; Richards, K. D.; Olsen, R. K. Tetrahedron Lett. 1976, 891.
- 24. Williams, D. H.; Fleming, I. Spectroscopic Methods in Organic Chemistry, 3<sup>rd</sup> ed., McGraw Hill Book Company: Maidenhead, U.K., 1980.
- 25. Poutsma, M. L. In Free Radicals, Kochi, J. K. Ed., Wiley: New York, 1973, vol. 2, p. 159.
- Kornblum, N.; Smiley, R. A.; Blackwood, R. K.; Iffland, D. C. J. Am. Chem. Soc. 1955, 77, 6269; Pocker, Y.; Kevill, D. N. J. Am. Chem. Soc. 1965, 87, 4760; Korblum, N.; Hardies, D. E. J. Am. Chem. Soc. 1966, 88, 1707.
- 27. Easton, C. J.; Hutton, C. A.; Tiekink, E. R. T. Z. Krist. 1993, 203, 310.
- 28. Bowden, K.; Heilbron, I. M.; Jones, E. R. H.; Weedon, B. C. L. J. Chem. Soc. 1946, 39.
- 29. March, J. Advanced Organic Chemistry, 3rd ed., John Wiley and Sons: New York, 1985, p. 220.
- 30. Sheehan, J. C.; Frank, V. S. J. Am. Chem. Soc. 1949, 71, 1856.
- 31. Marchand, J.; Pais, M.; Jarreau, F.-X. Bull. Soc. Chim. Fr. 1971, 3742.
- 32. Vogler, K. Helv. Chim. Acta 1950, 33, 2111.
- Sheehan, J. C.; Chapman, D. W.; Roth, R. W. J. Am. Chem. Soc. 1952, 74, 3822; Sheehan, J. C.; Goodman, M.; Hess, G. P. J. Am. Chem. Soc. 1956, 78, 1367; Clarke, S.; Hider, R. C.; John, D. I. J. Chem. Soc., Perkin Trans. 1 1973, 230; Hoogwater, D. A.; Reinhoundt, D. N.; Lie, T. S.; Gunneweg, J. J.; Beyerman, H. C. Recl. Trav. Chim. Pays-Bas. 1973, 92, 819; Hoffmann, E.; Schiff-Shenhav, H. J.Org. Chem. 1962, 27, 4686; Banks, G. R.; Cohen, D.; Pattenden, G. E.; Thomas, J. A. G. J. Chem Soc. (C) 1967, 126; Schwyzer, R.; Ludescher, U. Biochemistry 1968, 7, 2514; Greenlee, W. J.; Thorsett, E. D. J. Org. Chem. 1981, 46, 5351.

(Received in UK 4 March 1994; revised 18 April 1994; accepted 22 April 1994)